

Niðurstöður sívirkrar vöktunar á óæskilegum efnum í sjávarfangi úr auðlindinni 2020 /

Undesirable substances in seafood – results from the Icelandic marine monitoring activities in the year 2020

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Report summary



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Ágrip á íslensku:	_	20. Vöktunin hófst árið 200 verandi Atvinnuvega- og nvina gögnum og útgáfu á mabilinu 2003-2012. Vegnahlé á þessari mikilvægu ga 2013-2016. Verkefnið hófst á göngu yfir vöktun á óæskile em ætlað er til manneldis, e ekki lengur gerðar efnagrei u er að sýna fram á stöðu ís ýta gögnin við gerð áhætta og lýðheilsu. Verkefnið befna í efnahagslega mi	3 fyrir tilstuðlan þáverandi ýsköpunar-ráðneytisins, og skýrslum vegna þessarar á skorts á fjármagni í þetta agnasöfnun sem og útgáfu aftur í mars 2017 en vegna ægum efnum í ætum hluta en ekki fiskimjöl og lýsi fyrir iningar á PAH, PBDE og PFC slenskra sjávarafurða m.t.t. umats á matvælum til að
	endurskoðun er stöðugt na Almennt voru niðurstöðurr frá árunum 2003 til 2012 se sjávarafurðir innihalda óve varnarefni.	uðsynlegt. nar sem fengust 2020 í sam m og 2017 til 2019. Niðurst	nræmi við fyrri niðurstöður öðurnar sýndu að íslenskar
	í þessari skýrslu voru háma PCB (DL-PCB) og ekki díoxí nr. 1259/2011 notuð til að ESB. Niðurstöður ársins 202 vel undir hámarksgildum E styrkur svokallaðra ICES6-P hámarksgildi ESB samkva	nlík PCB (NDL-PCB) í matva i meta hvernig íslenskar sjá 20 sýna að öll sýni af sjávara SB fyrir þrávirk lífræn efni o CB efna vera lágur í ætum h æmt reglugerð nr. 1259, þungmálma, t.d. kadmíum	ávarafurðir standast kröfur afurðum til manneldis voru og þungmálma. Þá reyndist Iluta sjávarfangs, miðað við /2011. Sömuleiðis sýndu (Cd), blý (Pb) og kvikasilfur
Lykilorð á íslensku:	Sjávarfang, vöktun, Díoxín, hámarksgildi, heilnæmi, lýð	díoxínlík PCB, PCB, varnare	

Summary in English:

This report summarises the results obtained in 2020 for the screening of various undesirable substances in the edible part of Icelandic marine catches.

The main aim of this project is to gather data and evaluate the status of Icelandic seafood products in terms of undesirable substances and to utilise the data to estimate the exposure of consumers to these substances from Icelandic seafood and risks related to public health. The surveillance programme began in 2003 and was carried out for ten consecutive years before it was interrupted in 2013. The project was revived in March 2017 to fill in gaps of knowledge regarding the level of undesirable substances in economically important marine catches for Icelandic export. Due to financial limitations the surveillance now only covers screening for undesirable substances in the edible portion of marine catches for human consumption and not feed or feed components. The limited financial resources have also required the analysis of PAHs, PBDEs and PFCs to be excluded from the surveillance, providing somewhat more limited information than in 2013. However, it is considered a long-term project where extension and revision is constantly necessary.

In general, the results obtained in 2020 were in agreement with previous results on undesirable substances in the edible part of marine catches obtained in the monitoring years 2003 to 2012 and 2017 to 2019.

In this report from the surveillance programme, the maximum levels for dioxins, dioxin-like PCBs and non-dioxin-like PCBs in foodstuffs (Regulation No 1259/2011) were used to evaluate how Icelandic seafood products measure up to limits currently in effect.

The results show that in regard to the maximum levels set in the regulation, the edible parts of Icelandic seafood products contain negligible amounts of dioxins, dioxin like and non-dioxin-like PCBs. In fact, all samples of seafood analysed in 2020 were below EC maximum levels.

Furthermore, the concentration of ICES6-PCBs was found to be low in the edible part of the marine catches, compared to the maximum limits set by the EU (Commission Regulation 1259/2011).

The results also revealed that the concentrations of heavy metals, e.g. cadmium (Cd), lead (Pb) and mercury (Hg) in the edible part of marine catches were in all samples well below the maximum limits set by the EU.

English keywords:

Marine catches, monitoring, dioxin, PCB, pesticides, heavy metals, maximum limits, human consumption, public health

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1 Introduction

In 2003, the Icelandic Ministry of Fisheries, now the Ministry of Industries and Innovation, initiated a project aimed at screening for undesirable substances in the edible portion of marine catches, as well as in fish meal and fish oil for feed, captured in Icelandic waters. Matis was assigned the responsibility of carrying out the surveillance programme, which was ongoing for ten consecutive years. In the period 2013-2016 this important collection of information and publication of the results was interrupted since Matis did not receive funding to work on this monitoring project. However, in March 2017 the surveillance programme was revived with funding from the Ministry of Industries and Innovation in Iceland to gather data and evaluate the status of Icelandic seafood products regarding undesirable substances, however, the current funding only covers screening for undesirable substances in the edible portion of marine catches for human consumption not feed or feed components. The project includes measurements on various undesirable substances in several economically important marine species from Icelandic fishing grounds in order to gather information and evaluate the status of Icelandic seafood products in terms of undesirable substances. This report summarises results from the screening programme in the year 2020. The substances investigated in this monitoring project are: polychlorinated dibenzo dioxins and dibenzo furans (commonly called dioxins), dioxin-like polychlorinated biphenyls (PCBs), ICES-6 PCBs, 30 pesticides and breakdown products (i.e. HCB, DDTs, HCHs, dieldrin, endrin, chlordanes, toxaphenes and endosulfan substances), and inorganic trace elements such as heavy metals. The purpose of this work is:

- To gather information and evaluate the status of Icelandic seafood products in terms
 of undesirable substances.
- Provide scientific evidence that Icelandic seafood products conform to regulations on seafood safety. That is, to evaluate how products measure up to limits currently in effect for inorganic trace elements, organic contaminants and pesticides in the EU (Commission regulation (EC) No 1881/2006 and its amendments).
- 3. To utilise the data gathered in this programme for a risk assessment and the setting of maximum values within EU & the European Economic Area (EEA) area, which are constantly being reviewed based on new data.

4. Provide independent scientific data on undesirable substances in Icelandic seafood for food authorities, fisheries authorities, industry, markets and consumers.

In this report the maximum levels for dioxins, dioxin-like PCBs and non-dioxin-like PCBs in foodstuffs are used to evaluate how Icelandic seafood products measure up to European commission (EC) limits currently in effect. The results obtained in the years 2003 to 2012, as well as 2017 and 2018, have already been published and are accessible at the Matis website (http://www.matis.is: Auðunsson, 2004, Ásmundsdóttir et al., 2005, Ásmundsdóttir and Gunnlaugsdóttir, 2006, Ásmundsdóttir et al., 2008, Jörundsdóttir et al., 2009, Jörundsdóttir et al., 2010a, Jörundsdóttir et al., 2010b, Baldursdóttir et al., 2011, Jörundsdóttir et al., 2012, Jensen et al., 2013, Jensen et al., 2018, Jensen et al. 2019, Jensen et al. 2020). The above mentioned EU regulations have now been implemented (Reglugerð 265/2010) in the Icelandic legal framework regarding undesirable substances in food (Regulation (EC) No 1881/2006), which means that the maximum limits for undesirable substances in Icelandic seafood products are in line with the limits for these products in the EU member states.

2 Contaminants measured in the project

The following contaminants were measured in the edible parts of seafood and other seafood products for human consumption:

Dioxins, PCDD/Fs: Dioxins (dibenzo-p-dioxins) and dibensofurans (17 congeners according to WHO): 2.3.7.8-Tetra-CDD, 1.2.3.7.8-Penta-CDD, 1.2.3.4.7.8-Hexa-CDD, 1.2.3.6.7.8-Hexa-CDD, 1.2.3.7.8.9-Hexa-CDD, 1.2.3.4.6.7.8-Hepta-CDD, OCDD, 2.3.7.8-Tetra-CDF, 1.2.3.7.8-Penta-CDF, 2.3.4.7.8-Penta-CDF, 1.2.3.4.7.8-Hexa-CDF, 1.2.3.4.7.8-Hexa-CDF, 1.2.3.4.6.7.8-Hexa-CDF, 1.2.3.4.6.7.8-Hepta-CDF, 1.2.3.4.7.8-Hepta-CDF, 1.2.3.4.7.8-Hepta-CDF, 1.2.3.4.7.8-Hepta-CDF, OCDF.

Dioxin like PCB (12 congeners according to WHO):

non-ortho (CB-77, CB-81, CB-126, CB-169) and mono-ortho (CB-105, CB-114, CB-118, CB-123, CB-156, CB-157, CB-167, CB-189).

ICES-6-PCBs (6 congeners):

CB-28, CB-52, CB-101, CB-138, CB-153, CB-180.

Pesticides:

DDT-substances (6 congeners: pp-DDT, op-DDT, pp-DDD, op-DDD, pp-DDE and op-DDE), HCH-substances (4 isomers: α -, β -, γ -(Lindane), and δ -hexachlorocyclohexan), HCB, chlordanes (4 congeners and isomers: α - and γ -chlordane, oxychlordane and trans-nonachlor), toxaphenes (3 congeners, P 26, 50 and 62), aldrin, dieldrin, endrin, endosulfan (3 congeners and isomers: α - and β -endosulfan and endosulfansulfat) and heptachlor (3 congeners: heptachlor, cishepatchlorepoxid, trans-heptachlorepoxid).

Inorganic trace elements:

Hg (mercury), Cd (cadmium), Pb (lead), total As (organic and inorganic arsenic), chromium (Cr) and tin (Sn).

3 Sampling and analysis

3.1 Sampling

The collection of samples and the quality criteria for the analytical methods were in accordance with conditions set out by the EU for the information gathering campaign on dioxins and dioxin-like PCBs as well as for metals (Commission regulation 333/2007/EC, Commission regulation 2017/644/EC). The fish samples were collected by the Marine and Freshwater Research Institute (MRI) in Iceland according to sampling protocols provided by Matis and the samples were kept frozen until preparation for analysis (see section 4.1.1). The blue mussels were taken at the production site prior to any production and thus represent the raw material from this fishing ground. The sea urchin roe was provided by the producer, while the cod livers were taken at the production site as soon as the fish had been gutted – in both cases care was taken to avoid any cross contamination from the production environment. Fishing grounds around Iceland are divided into five areas, as illustrated in Figure 1. All the samples were identified and labelled with the fishing area where they were caught.

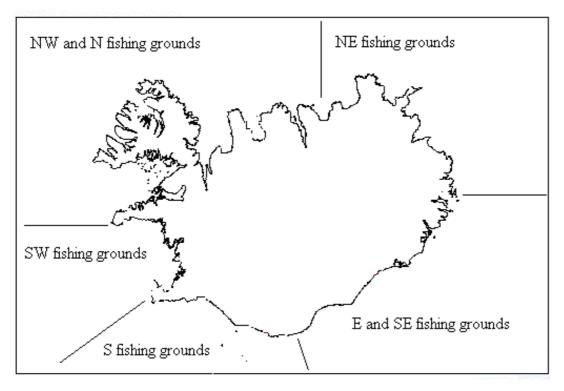


Figure 1: The division of the fishing grounds around Iceland used in this research.

3.1.1 Sample preparation

All analyses were performed on the edible parts of the fish, blue mussels, sea urchin roes and cod liver samples. Each fish sample consisted of a pool from at least ten individuals of a specific length distribution. For details on length distribution and fishing grounds of the samples see Table 1 and 2 in the Appendix. Prior to sample preparation each fish was defrosted, after that the total weight and length of each individual fish was recorded as well as gender, gut weight and weight of fillets. The skinless fish fillets from the individuals were then pooled, homogenised and frozen again for analysis of organic contaminants or freeze-dried for heavy metal analysis. The cod liver sample consisted of a pooled sample from 10 individuals, the cod liver was removed from the cod soon after it has been gutted at the production site and put in a clean container. These cod livers were delivered fresh to the laboratory, where they were homogenised and pooled to prepare one composite sample. The blue mussels were delivered frozen by the producer to the laboratory were the shell was removed and the edible flesh of around 1 kg pooled and homogenised in a food blender prior to analysis. Approximately 500 g of sea urchin roe was delivered frozen to the laboratory where it was thawed, homogenised and pooled to prepare one composite sample.

3.2 Analyses

The heavy metal analysis of chromium, arsenic, tin, cadmium, mercury and lead was carried out at Matís. Inorganic contaminants in samples were determined by ICP-MS according to an accredited in-house method SV-25-02-SN in Matis Quality manual (modified NMKL 186 (2007) method). Matís is a National Reference Laboratory for heavy metal analysis in food and feed and has been taking part in various international inter-laboratory studies for many years.

The lipid content and organic contaminants were measured by Eurofins, Hamburg, Germany. Eurofins has taken part in an international inter-laboratory quality control study organised by WHO and EU and uses accredited methods for analysing lipids, dioxin, WHO-PCBs, ICES-6-PCBs, and pesticides.

All results are expressed as upper bond level, which means that when the concentration of a substance is measured to be below limit of detection (LOD) or limit of quantification (LOQ) of the analytical method, the concentration is set to be equal to the LOD/LOQ. In the case of dioxins and dioxin-like PCBs, the analytical data are converted to pg/g WHO-TEQ where the toxicity of each congener has been calculated using WHO-TEF (Toxic Equivalence Factor) based on the existing knowledge of its toxicity (Van den Berg et al., 1998). WHO-TEQ values have been adapted by the World Health Organization (WHO) in 1997 and by EU in its legislations. In 2005 the WHO-TEF values were re-evaluated based on existing toxicological data (Van den Berg et al., 2005, Haws et al., 2006) and expert judgment. These new TEF values have been established as the WHO-2005-TEQs for human risk assessment of the concerned compounds and have been implemented in the current EU legislation i.e. Commission Regulation (EU) No 1259/2011.

4 Results from monitoring of marine catches in Iceland

All results for undesirable substances from the surveillance programme in 2020 are listed in Tables 1-3 in the Appendix. The sections below contain an overview of the results obtained in samples of fish and other seafood products taken as part of the monitoring activities 2020.

4.1 Dioxins (PCDD/Fs) and dioxin like PCBs

4.1.1 Dioxins and dioxin like PCBs in seafood

All the species analysed contained dioxins (PCDD/PCDFs) below EU maximum limits (Figure 2 and Table 1 in the Appendix).

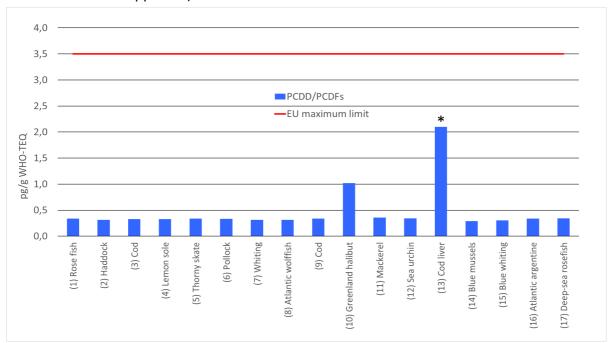


Figure 2: Dioxins (PCDD/PCDFs) in the edible part of marine catches from Icelandic fishing grounds in 2020 in relation to maximum EU limit in WHO-TEQ pg/g wet weight. The number within parenthesis is the sample number indicated in Table 1 in the Appendix.

As in previous years, a small difference was observed in the dioxin content between different marine species. The species that accumulate fat in the muscle, like Greenland halibut (sample no. 10), contain more dioxins than species which accumulate fat in the liver and thus have almost no fat in the muscle. This can be clearly seen when comparing the cod samples (samples no. 3 and 9), which are skinless cod fillets, with the cod livers (sample no. 13). In this case, the dioxin content in the cod muscle was 0,33 pg/g WHO-TEQ compared to 2,1 pg/g WHO-TEQ in the cod liver.

^{*}EU maximum limit for fish liver not established.

In general, the level of dioxins in the edible part of the fish increases as the fat percentage in the muscle increases, but other important variables are age (size) and habitat. Greenland halibut can become quite old, which probably contributes considerably to the higher dioxins and dioxin-like PCBs values observed for this species, while Mackerel is high in fat content but does not become very old and therefore accumulates less dioxins over his lifetime (Table 1 in the Appendix). Figure 3 shows the sum of dioxins and dioxin-like PCBs in all samples analysed. All the species analysed contained total dioxins and dioxin-like PCBs below EU maximum limits (Figure 3 and Table 1 in the Appendix).

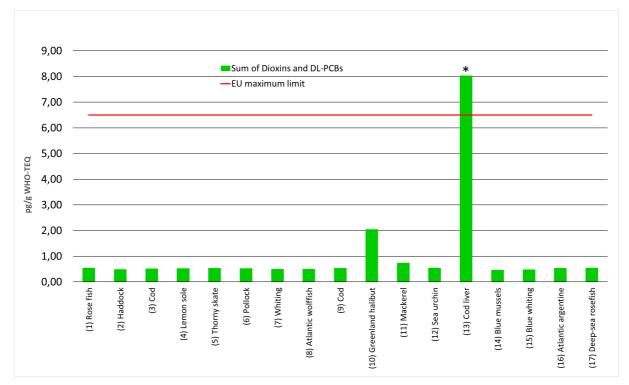


Figure 3: Sum of dioxins and dioxin-like PCBs in the edible part of marine catches from Icelandic fishing grounds in 2020 in relation to maximum EU limit in WHO-TEQ pg/g wet weight. The number within parenthesis is the sample number indicated in Table 1 in the Appendix.

The results show that the sum of dioxins and dioxin-like PCBs in the cod liver was considerably higher than in the muscle samples of the marine species analysed. However, the maximum limit set by the EU for this product is also considerably higher i.e. 20 pg/g WHO-TEQ wet weight compared to 6,5 pg/g WHO-TEQ wet weight for fish muscle and crustaceans. Therefore, the observed value, 8,03 pg/g WHO-TEQ for the cod liver is well below the limit set by the EU.

^{*}EU maximum limit for fish liver is 20 pg/g wet weight.

4.2 Marker PCBs

Marker PCBs have been used as indicators of the total PCB content or body burden of environmental biota, food and human tissue. The most frequent approach is to use either the total level of six or seven of the most commonly occurring PCBs. Nevertheless, the EU maximum limits are set for the sum concentration of ICES-6, i.e. CB-28, -52, -101, -138, -153 and -180 (Commission Regulation (EU) No 1259/2011). To enable comparison to earlier results, the sum of seven marker PCBs are presented in Table 1 in the Appendix, while the ICES-6 maximum limits are presented in Figure 4 to evaluate how Icelandic seafood products measure up to EU maximum limits.

4.2.1 ICES-6 PCBs in seafood

The results obtained for all of the Icelandic marine catches were well below the maximum limit set for non-dioxin-like PCBs i.e. the so-called ICES-6 (Figure 4).

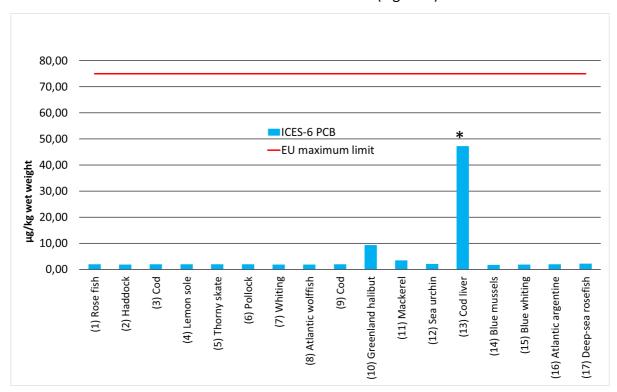


Figure 4: ICES-6 PCBs in marine catches from Iceland in 2020 (in μ g/kg wet weight). Number in parenthesis is the sample number designated to each sample, see Table 1 in Appendix.

In this study, the highest total concentration for <u>the sum</u> of all six marker PCBs in the muscle samples was measured in Greenland halibut (sample no. 10, Figure 4), a total of 9,2 μ g/kg wet

^{*}EU maximum limit for fish liver is 200 μ g/kg wet weight.

weight, the highest individual PCB congener measured in the Greenland halibut was PCB-153 with 3,34 μ g/kg wet weight, or approximately one third of the total. As for the dioxins and dioxin-like PCBs (section 4.1.1.), the highest concentrations of the ICES-6 PCBs were found in species with higher lipid content in the muscle. Likewise, the ICES-6 PCB concentration was considerably higher in the cod liver sample than in the muscle samples analysed (Figure 4 and Table 1 in the Appendix). However, the EU maximum limit is also considerably higher (200 μ g/kg wet weight) for fish liver than for the muscle meat of the other samples investigated. For details, see Table 1 in the Appendix.

4.3 Polycyclic aromatic hydrocarbons (PAHs)

PAHs were not analysed in the samples this year. Results on PAHs in Icelandic seafood have been published in previous reports (Jörundsdóttir et al., 2010, Jensen et al., 2013).

4.4 Brominated flame retardants (BFRs)

BFRs have been accumulating in the environment over the last decade as their use in industry has increased. BFRs were not analysed in the samples this year. Results on BFRs in Icelandic seafood have been published in previous reports (Jensen et al., 2013).

4.5 Pesticides

In this section, the results for 12 different classes of pesticides are discussed. Results are shown in Table 2 in the Appendix.

In total 12 different pesticides or groups of pesticides were measured in the monitoring programme.

DDT (dichloro diphenyl trichloroethan) is probably the best-known insecticide. The technical product DDT is fundamentally composed of p,p'-DDT (80%) (Buser, 1995). DDT breaks down in nature, mostly to DDE but also to DDD. The concentration of DDT presented in this report is the sum of p,p'-DDT, o,p'-DDT, p,p'-DDE, o,p'-DDE, p,p'-DDD and o,p'-DDD.

HCH (hexachlorocyclohexan) is an insecticide which has been used since 1949. It is still produced and used in numerous countries, although it has been banned in many countries since the 1970s. Technical-grade HCH is a mixture of mainly four isomers: α -, β -, γ -(Lindane), and δ -HCH. Of these, only Lindane is an active substance comprising of approximately 15% of

the total mixture, while α -HCH is 60-70% of the mixture. The Food and Agriculture Organization of the UN (FAO) has prohibited the use of the HCH mixture since in the 1980s, after that it was only allowed to use 99% pure Lindane. In this report the concentration of α -, β -, γ -(Lindane), and δ -HCH in the samples are reported.

HCB (hexachlorobenzene) is a fungicide, but it has also been used for industrial purpose and was e.g. produced in Germany until 1993. Today, HCB is mainly a by-product in different industrial processes such as production of pesticides but also from waste incineration and energy production from fossil fuel.

Chlordanes is a group of compounds and isomers where α - and γ -chlordane, oxychlordane and *trans*-nonachlor are the most common, but over 140 different chlordanes were produced from 1946 until 1988 when the production was banned. Chlordanes have been widely used all over the world as insecticides. In this report the concentration of chlordanes is reported as the sum of α -chlordane, γ -chlordane and oxychlordane. *Trans*-nonachlor is reported separately.

The **Toxaphenes** measured in the samples are the so-called parlar 26, 50 and 62. Toxaphene was used as an insecticide after the use of DDT was discontinued. Toxaphenes use was widespread and the toxaphene congeners are numerous. Several hundred have been analysed but they are thought to be tens of thousands. The substances measured, i.e. the parlar 26, 50 and 62, are the most common toxaphenes (about 25% of the total amount in nature) and these are used as indicators of toxaphene pollution. In this report the concentration of toxaphenes is reported as the sum of toxaphene 26, 50 and 62.

Aldrin and Dieldrin are widely used insecticides, but in plants and animals aldrin is transformed to dieldrin. Hence, the concentration of aldrin was below LOD in all the samples measured, while dieldrin was in some samples above LOD. The results are presented as the sum of these two.

Two **Endosulfans** were measured, α - and β -endosulfan, as well as endosulfansulfat which is the breakdown product of endosulfan. Endosulfans are not as persistent as the other insecticides measured in this project. In this report the concentration of endosulfans is reported as the sum of α -endusulfan, β -endusulfan and endosulfansulfat. Other pesticides measured were **Endrin**, the sum of **Heptachlores** (cis-heptachlorepoxide, transheptachlorepoxide and heptachlor), **Pentachlorobenzene**, **Mirex** and **Octachlorostyrene**.

4.5.1 Pesticides in seafood

The results show that most of the pesticides measured in marine catches from Icelandic waters were below the limit of detection (see Table 2 in the Appendix). However, as mentioned before the results are expressed as upper bond, and therefore the results presented are likely to be an overestimation. HCB and *trans*-Nonachlor were detected in about half of the species analysed, whilst α - β -, γ - and δ -HCHs were almost always below LOQ. Figure 5 shows the level of total DDT in the different marine catches, while Figure 6 shows the level of HCB in the same samples.

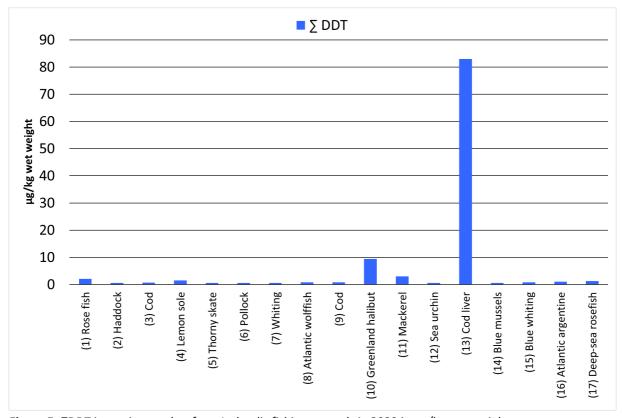


Figure 5: Σ DDT in marine catches from Icelandic fishing grounds in 2020 in μ g/kg wet weight.

No limits have yet been set for pesticides in seafood, but to enable comparison with earlier measurements presented in previous reports from this project (Jensen, et al. 2013, Jensen, et al. 2018, Jensen, et al. 2019 & Jensen, et al. 2020), the results of Σ DDT and HCB are presented from the monitoring in 2020. In general, the concentration of pesticides is higher in Icelandic marine catches with a higher lipid content (Table 2 in Appendix).

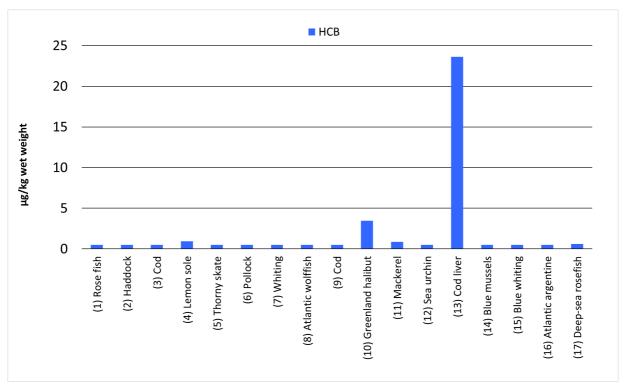


Figure 6: HCB in marine catches from Icelandic fishing grounds in 2020 in $\mu g/kg$ wet weight.

4.6 Inorganic trace elements

Inorganic trace elements were analysed in all samples from the year 2020. The following inorganic trace elements were analysed: Hg (mercury), Cd (cadmium), Pb (lead), As (arsenic), Sn (tin) and Cr (chromium). As mentioned before, the results are expressed as upper bond and therefore the results presented are likely to be an overestimation. All results for the analysed trace elements are reported in Table 3 in the Appendix.

4.6.1 Inorganic trace elements in seafood

In short, the concentration of the heavy metal Hg, Pb and Cd in all samples consisting of the edible part of fish and the cod liver were well below the maximum limits set by EU (Commission regulation (EC) No 1881/2006 and its amendments). Maximum limits set by the EU (Commission regulation 1881/2006) for tin (Sn) only apply to canned food products and no maximum limits exist in the EU for tin (Sn) in fish or fishery products. The concentration of tin (Sn) in all the samples analysed was very low as can be seen in Table 3 in the Appendix; in fact, no sample contained tin in concentrations above limits of detection.

The concentration of mercury (Hg) in the marine catches is shown in Figure 7.

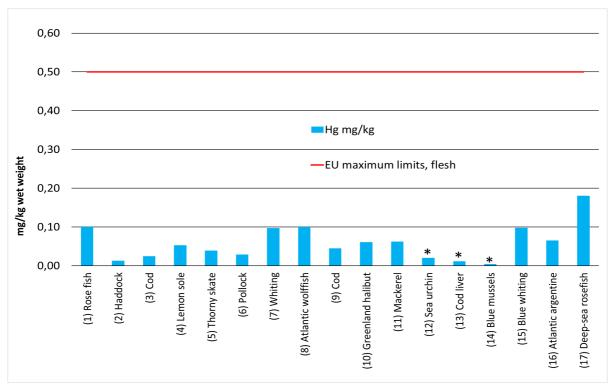


Figure 7: Mercury (Hg) in marine catches from Icelandic fishing grounds in 2020 in mg/kg wet weight. *EU maximum limit not established.

The concentration of lead (Pb) in all the samples of marine catches was very low as can be seen in Figure 8 and Table 3 in the Appendix; in fact, the concentration of lead was below the limit of detection for all samples.

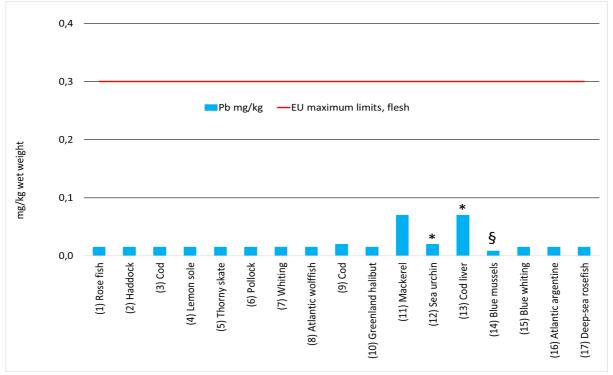


Figure 8: Lead (Pb) in marine catches from Icelandic fishing grounds in 2020 in mg/kg wet weight.

*EU maximum limit for Pb does not apply to cod liver or sea urchin since it is set for muscle meat of fish.

§EU maximum limit for Pb in bivalve molluscs is set to 1,5 mg/kg wet weight

The concentration of cadmium (Cd) in the marine catches was generally very low as can be seen in Figure 9 and Table 3 in the Appendix. No sample contained cadmium in concentration above EU maximum limits. The EU maximum limit for cadmium in bivalve molluscs is set to 1,0 mg/kg wet weight. The EU maximum limit for Cd shown in Figure 9 does neither apply to cod liver nor sea urchin since it is set for muscle meat of fish and is therefore not applicable for cod liver or sea urchin.

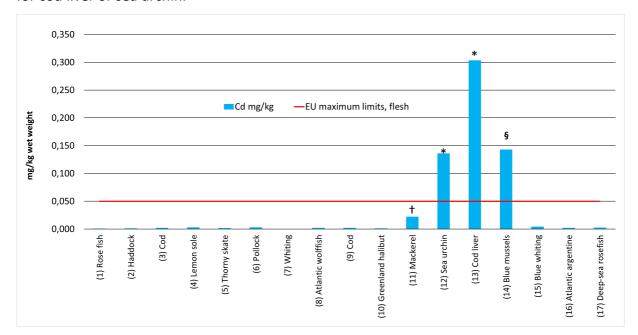


Figure 9: Cadmium (Cd) in marine catches from Icelandic fishing grounds in 2020 in mg/kg wet weight. †EU maximum limit for Cd in mackerel is set to 0,1 mg/kg wet weight.

No limits have yet been set for arsenic in foodstuffs, but results from the monitoring in 2020, which are shown in Figure 10 were mostly in agreement with earlier measurements (Auðunsson, 2004, Ásmundsdóttir et al. 2005, Ásmundsdóttir and Gunnlaugsdóttir, 2006, Jörundsdóttir et al., 2009, Baldursdóttir et al., 2011, Jörundsdóttir et al., 2012, Jensen et al., 2013, Jensen et al., 2018, Jensen et al., 2019 & Jensen, et al. 2020). The highest levels of As (above 10 mg/kg) were found in the lemon sole sample (sample no. 4) and the thorny skate (sample no. 5) as seen in Figure 10. This is different from the results from 2019 (Jensen et al., 2020) where all samples contained arsenic below 10 mg/kg. The total arsenic concentration was measured in the samples, but not the concentration of the toxic form i.e. inorganic arsenic.

[§] EU maximum limit for Cd in bivalve molluscs is set to 1,0 mg/kg wet weight.

^{*}EU maximum limit for Cd does not apply to cod liver or sea urchin since it is set for muscle meat of fish.

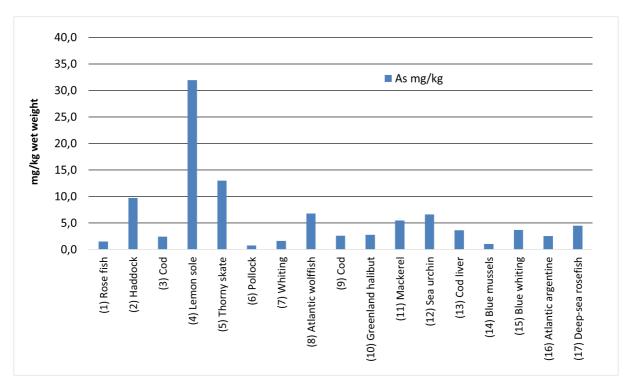


Figure 10: Arsenic (As) in marine catches from Icelandic fishing grounds in 2020 in mg/kg wet weight.

5 Acknowledgements

Special thanks to the Icelandic marine and freshwater research institute (MRI) for the sampling of all fish samples. Thanks to Royal Iceland for providing the sea urchin roe, Akraborg for the samples of cod livers and Neskel for providing blue mussels for this project.

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7 Appendix

Sample code FNN Sample mame Latin name Triving Size Lipid content PCDD/PCDFs Dioan like PCBs Di-ACBs R20-1256-1 2.1 Rose fish Schoates marrinus SW 40-49 % pgg WHO-TFQ	Rose 2	Sample name	_								
Schastes marinus Schastes marinus % pag/g WHO-TEQ pg/g WHO-TEQ Relanogrammus aeglefinus SW 30-40 3 0,34 0,21 Gadus morhua N 40-49 0,6 0,33 0,20 Microstomus aeglefinus N 50-59 0,6 0,33 0,20 Microstomus N 40-60 0,6 0,33 0,20 Raja Ambyraja radiata NE 40-60 0,6 0,33 0,20 Pollachius virens NW 60-70 0,8 0,33 0,20 Merlangius merlangus SE 40-60 0,6 0,31 0,19 Anarhichas lupus SE 40-60 0,6 0,31 0,19 Anarhichas lupus N 60-70 0,8 0,31 0,19 Anarhichas lupus N 60-70 0,8 0,34 0,20 Scomber scombrus N 50-60 11,4 1,02 0,18 Argentina silus N N N	- 2 % 4 % 9	fig.h	Latin name	FISHING		Lipid content	PCDD/PCDFs	Dioxin like PCBs	DL-PCBs	PCBs	PCB
Sebastes marinus SW 30-40 3 0,34 0,21 Melanogrammus aeglefinus NE 40-49 0,6 0,31 0,19 Gadus morhua N 50-59 0,6 0,33 0,20 Raja Amblyraja radiata NE 40-60 0,6 0,34 0,20 Pollachius virens NE 40-60 0,6 0,34 0,20 Pollachius virens NW 60-70 0,8 0,34 0,20 Marilachius virens NW 60-70 0,8 0,31 0,19 Marilachius virus NW 60-70 0,6 0,31 0,19 Reinhardtius hippoglossoides N 60-70 0,8 0,34 0,20 Reinhardtius hippoglossoides N 50-60 11,4 1,02 1,03 Scomber scombrus NW n.a 5,0 0,34 0,21 Adus morhua n.a 7,0 2,10 2,93 Myrilus edulis NW 0,6 0,34	- 2 % 4 % 9 t	fich		Si onin	[cm]	%	pg/g WHO-TEQ	pg/g WHO-TEQ	pg/g WHO-TEQ	µg/kg	µg/kg
Metanogrammus aeglefinus NE 40-49 0,6 0,31 0,19 Gadus morhua N 50-59 0,6 0,33 0,20 Microstomus kitt NE 40-60 0,6 0,34 0,20 Pollachius virens NE 40-60 0,6 0,34 0,20 Pollachius virens NW 60-70 0,8 0,31 0,19 Merlangius merlangus SE 30-40 0,6 0,31 0,19 Anerlangus merlangus SE 30-40 0,6 0,31 0,19 Anerlangus merlangus N 60-70 0,8 0,34 0,20 Reinhardius hippoglossoides N 50-60 11,4 1,02 1,03 Scomber scombrus n.a 30-40 26,5 0,34 0,20 Areinhoidea NW n.a 5,0 0,34 0,21 Myrilus edulis NW n.a 0,6 0,3 0,34 Argentina silus SE 40-50 <t< td=""><td>0 w 4 w 0 t</td><td>11911</td><td>Sebastes marinus</td><td>MS</td><td>30-40</td><td>3</td><td>0,34</td><td>0,21</td><td>0,54</td><td>2,05</td><td>1,97</td></t<>	0 w 4 w 0 t	11911	Sebastes marinus	MS	30-40	3	0,34	0,21	0,54	2,05	1,97
Gadus morhua N 50-59 0,6 0,33 0,20 Microstomus kitt NE 30-50 0,6 0,33 0,20 Raja Amblyraja radiata NE 40-60 0,6 0,34 0,20 Pollachius virens NW 60-70 0,8 0,33 0,20 Merlangius merlangus NW 60-70 0,6 0,31 0,19 Anathichas lupus SE 40-50 0,6 0,31 0,19 Anathichas lupus N 60-70 0,8 0,31 0,19 Anathichas lupus N 60-70 0,6 0,31 0,19 Reinharditus hippoglossoides N 50-60 11,4 1,02 1,03 Scomber scombrus NW n.a 30-40 26,5 0,36 0,38 Gadus morhua NW n.a 67,0 2,10 2,93 Mytilus edulis NW n.a 0,6 0,20 0,18 Argentina silus SE 40-50 <td>w 4 w 0 i</td> <td>ck</td> <td>Melanogrammus aeglefinus</td> <td>NE</td> <td>40-49</td> <td>9,0</td> <td>0,31</td> <td>0,19</td> <td>0,50</td> <td>1,85</td> <td>1,81</td>	w 4 w 0 i	ck	Melanogrammus aeglefinus	NE	40-49	9,0	0,31	0,19	0,50	1,85	1,81
Raja Amblyraja radiata NE 30-50 0,6 0,33 0,20 Pollachius virens NE 40-60 0,6 0,34 0,20 Pollachius virens NW 60-70 0,8 0,33 0,20 Merlangius merlangus SE 40-50 0,6 0,31 0,19 Anarhichas lupus N 60-70 0,8 0,31 0,19 Gadus morhua N 60-70 0,8 0,34 0,19 Reinhardius hippoglossoides N 50-60 11,4 1,02 1,03 Scomber scombrus N 50-60 11,4 1,02 0,38 Echinoidea NW n.a 30-40 26,5 0,36 0,38 Gadus morhua n.a n.a 67,0 2,10 5,93 Mytilus edulis NW 1.2 0,54 0,18 Argentina silus SE 40-50 2,3 0,34 0,21 Sebastes mentella SW 40-50 2,3	4 % 0 1		Gadus morhua	N	50-59	9,0	0,33	0,20	0,52	1,94	1,90
Raja Amblyraja radiata NE 40-60 0,6 0,34 0,20 Pollachius virens NW 60-70 0,8 0,33 0,20 Merlangius merlangus SE 40-50 0,6 0,31 0,19 Anarhichas lupus SE 30-40 0,6 0,31 0,19 Radus morhua N 60-70 0,8 0,34 0,20 Reinhardtius hippoglossoides N 50-60 11,4 1,02 1,03 Scomber scombrus N 50-60 11,4 1,02 1,03 Scomber scombrus NW n.a 30-40 26,5 0,36 0,38 Echinoidea NW n.a 67,0 2,10 5,93 Mytilus edulis NW n.a 0,6 0,29 0,18 Argentina silus SE 40-50 1,3 0,34 0,21 Sebastes mentella SW 40-50 2,3 0,34 0,21 Sebastes mentella SW 40-50 </td <td>W O 1</td> <td>ı sole</td> <td>Microstomus kitt</td> <td>NE</td> <td>30-20</td> <td>9,0</td> <td>0,33</td> <td>0,20</td> <td>0,53</td> <td>1,95</td> <td>1,91</td>	W O 1	ı sole	Microstomus kitt	NE	30-20	9,0	0,33	0,20	0,53	1,95	1,91
Pollachius virens NW 60-70 0,8 0,33 0,20 Merlangius merlangus SE 40-50 0,6 0,31 0,19 Anarhichas lupus SE 30-40 0,6 0,31 0,19 Gadus morhua N 60-70 0,8 0,34 0,20 Reinhardtius hippoglossoides N 50-60 11,4 1,02 1,03 Scomber scombrus N 50-60 11,4 1,02 1,03 Scomber scombrus NW n.a 30-40 26,5 0,36 0,38 Echinoidea NW n.a 67,0 2,10 5,93 Myillus edulis NW n.a 67,0 2,10 5,93 Myillus edulis NW n.a 67,0 0,29 0,18 Argentina silus SE 40-50 2,3 0,34 0,21 Sebastes mentella NW 40-50 2,3 0,34 0,21 S.50 A 40-50 2,3 <td>91</td> <td>y skate</td> <td>Raja Amblyraja radiata</td> <td>NE</td> <td>40-60</td> <td>9,0</td> <td>0,34</td> <td>0,20</td> <td>0,54</td> <td>2,01</td> <td>1,96</td>	91	y skate	Raja Amblyraja radiata	NE	40-60	9,0	0,34	0,20	0,54	2,01	1,96
Merlangus merlangus SE 40-50 0,6 0,31 0,19 Anarhichas lupus SE 30-40 0,6 0,31 0,19 Gadus morhua N 60-70 0,8 0,34 0,20 Reinhardius hippoglossoides N 50-60 11,4 1,02 1,03 Scomber scombrus N 50-60 11,4 1,02 1,03 Echinoidea NW n.a 5,0 0,36 0,38 Ayilus edulis NW n.a 67,0 2,10 5,93 Myilus edulis NW n.a 67,0 2,10 5,93 Myilus edulis NW n.a 67,0 2,10 5,93 Argentina silus SE 40-50 1,2 0,34 0,18 Sebastes mentella SW 40-50 2,3 0,34 0,21 Sebastes mentella SS 40-50 2,3 0,34 0,21 3,-156,-157,-167 and -189 3,50 * *	t	k	Pollachius virens	NW	02-09	8,0	0,33	0,20	0,53	1,99	1,94
Anarhichas lupus SE 30-40 0,6 0,31 0,19 Gadus morhua N 60-70 0,8 0,34 0,20 Reinhardius hippoglossoides N 50-60 11,4 1,02 1,03 Scomber scombrus n.a 30-40 26,5 0,36 0,38 0,20 Echinoidea NW n.a 5,0 0,34 0,21 3,83 Myvilus edulis NW n.a 67,0 2,10 5,93 Myvilus edulis NW n.a 67,0 2,10 5,93 Myvilus edulis NW n.a 67,0 0,21 0,21 Argentina silus SE 40-50 1,2 0,34 0,21 Sebastes mentella SW 40-50 2,3 0,34 0,21 3,-156,-157,-167 and -189 3,50 * *		ng		SE	40-50	9,0	0,31	0,19	0,50	1,85	1,81
Gadus morhua N 60-70 0,8 0,34 0,20 Reinhardtius hippoglossoides N 50-60 11,4 1,02 1,03 Scomber scombrus n.a 30-40 26,5 0,36 0,38 Echinoidea NW n.a 5,0 0,34 0,21 Gadus morhua n.a n.a 67,0 2,10 5,93 Myrilus edulis NW n.a 67,0 0,29 0,18 Micromesistius poutassou NW 30-40 1,2 0,39 0,18 Argentina silus SE 40-50 1,3 0,34 0,20 Sebastes mentella SW 40-50 2,3 0,34 0,21 3,-156,-157,-167 and -189 3,50 * *	∞	tic wolffish	Anarhichas Iupus	SE	30-40	9,0	0,31	0,19	0,50	1,85	1,81
Reinhardtius hippoglossoides N 50-60 11,4 1,02 1,03 Scomber scombrus n.a 30-40 26,5 0,36 0,38 Echinoidea NW n.a 5,0 0,34 0,21 Gadus morbua n.a n.a 67,0 2,10 5,93 Myrilus edulis NW n.a 0,6 0,29 0,18 Micromexistius poutassou NW 30-40 1,2 0,39 0,18 Argentina silus SE 40-50 1,3 0,34 0,20 Sebastes mentella SW 40-50 2,3 0,34 0,20 3,-156,-157,-167 and -189 3,50 * *	6		Gadus morhua	N	02-09	8,0	0,34	0,20	0,54	2,00	1,95
Scomber scombrus n.a 30-40 26,5 0,36 0,38 Echinoidea NW n.a 5,0 0,34 0,21 Gadus morhua n.a 67,0 2,10 5,93 Myrilus edulis NW n.a 0,6 0,29 0,18 Micromesistius poutassou NW 30-40 1,2 0,30 0,18 Argentina silus SE 40-50 1,3 0,34 0,20 Sebastes mentella SW 40-50 2,3 0,34 0,20 3,-156,-157,-167 and -189 3,50 * *	10	land halibut	Reinhardtius hippoglossoides	N	20-60	11,4	1,02	1,03	2,05	10,8	9,24
Echinoidea NW n.a 5,0 0,34 0,21 Gadus morhua n.a n.a 67,0 2,10 5,93 Myilus edulis NW n.a 0,6 0,29 0,18 Micromesistius poutassou NW 30-40 1,2 0,30 0,18 Argentina silus SE 40-50 1,3 0,34 0,20 Sebastes mentella SW 40-50 2,3 0,34 0,20 3,-156,-157,-167 and -189 3,50 *	11	erel	Scomber scombrus	n.a	30-40	26,5	0,36	0,38	0,74	3,87	3,4
Gadus morhua n.a n.a 67,0 2,10 5,93 Myilus edulis NW n.a 0,6 0,29 0,18 Micromesistius poutassou NW 30-40 1,2 0,30 0,18 Argentina silus SE 40-50 1,3 0,34 0,20 Sebastes mentella SW 40-50 2,3 0,34 0,20 3,-156,-157,-167 and -189 3,50 * *	12	chin	Echinoidea	NW	n.a	5,0	0,34	0,21	0,55	2,04	1,98
Myitlus edulis N.W n.a 0,6 0,29 0,18 Micromesistius poutassou N.W 30-40 1,2 0,30 0,18 Argentina silus SE 40-50 1,3 0,34 0,20 Sebastes mentella SW 40-50 2,3 0,34 0,20 3,-156,-157,-167 and -189 3,50 * *	13	ver	Gadus morhua	n.a	n.a	67,0	2,10	5,93	8,03	55,6	47,2
Micromesistius poutassou NW 30-40 1,2 0,30 0,18 Argentina silus SE 40-50 1,3 0,34 0,20 Sebastes mentella SW 40-50 2,3 0,34 0,21 3,-156, -157, -167 and -189 3,50 * *	14	unssels	Mytilus edulis	NW	n.a	9,0	0,29	0,18	0,47	1,74	1,69
Argentina silus SE 40-50 1,3 0,34 0,20 0 Sebastes mentella SIV 40-50 2,3 0,34 0,21 0 3,50 * 3,50 * 0 3,-156, -157, -167 and -189 0,21 0 0	15	vhiting	Micromesistius poutassou	NW	30-40	1,2	0,30	0,18	0,48	1,79	1,75
Sebastes mentella SW 40-50 2,3 0,34 0,21 0,2	16	tic argentine	Argentina silus	SE	40-50	1,3	0,34	0,20	0,54	2,00	1,95
3,-156,-157,-167 and -189	17		Sebastes mentella	SW	40-50	2,3	0,34	0,21	0,55	2,29	2,10
3,-156,-157,-167 and -1	EU ma:	vximum limits‡					3,50	*	6,50	*	75
2,3,7,5,8-PCDDs and PCDFs. 77, -81, -126, -169, -105, -114, -118, -123, -156, -157, -167 and -1	*No maximum limits exist in the EU fo	or the substances									
118, -123, -156, -157, -167 and -1	n.a. Not available	J PCDE.									
	PCBs are CB-77 -81 -126 -149 -11	id FCDFs. 05 -114 -118 -123 -1									
Marker PCBs are CB-28 -52 -101 -118 -138 -153 and -180	Warker PCBs are CB-28 -52 -101 -118	8 -138 -153 and -180	, 101, 101 min								

Table 2: Pest	cides in marii.	re catches and one cod I	laine 2: Festicides in marme catches and one cod liver sample on wet weight										=	
	Fish sample		:	Fishing	Size	Lipid					i i	Pentachlor		M
Sample code	no.	Sample name	Latin name	ground	[cm]	content %	a-HCH mg/kg	P-HCH mg/kg	G-HCH mg/kg	g-HCH mg/kg	2 DD1 mg/kg	benzene mg/kg	HCB mg/kg	Heptachiores mg/kg
R20-1226-1	1	Rose fish	Sebastes marinus	MS	30-40	3	<0,25	<0,25	<0,25	<0,25	2,1	<0,50	0,50	0,55
R20-1226-2	2	Haddock	Melanogrammus aeglefinus	NE	40-49	9,0	<0,25	<0,25	<0,25	<0,25	9,0	<0,50	<0,50	0,55
R20-1226-3	3	Cod	Gadus morhua	N	50-59	9,0	<0,25	<0,25	<0,25	<0,25	0,7	<0,50	<0,50	0,55
R20-1226-4	4	Lemon sole	Microstomus kitt	NE	30-50	9,0	<0,46	<0,46	<0,46	<0,46	1,5	<0,93	<0,93	1,02
R20-1226-5	5	Thomy skate	Raja Amblyraja radiata	NE	40-60	9,0	<0,25	<0,25	<0,25	<0,25	9,0	<0,50	<0,50	0,55
R20-1226-6	9	Pollock	Pollachius virens	MN	02-09	8,0	<0,25	<0,25	<0,25	<0,25	9,0	<0,50	<0,50	0,55
R20-1226-7	7	Whiting	Merlangius merlangus	SE	40-50	9,0	<0,25	<0,25	<0,25	<0,25	9,0	<0,50	<0,50	0,55
R20-1226-8	∞	Atlantic wolffish	Anarhichas lupus	SE	30-40	9,0	<0,25	<0,25	<0,25	<0,25	8,0	<0,50	<0,50	0,55
R20-1226-9	6	Cod	Gadus morhua	N	02-09	8,0	<0,25	<0,25	<0,25	<0,25	8,0	<0,50	<0,50	0,55
R20-1226-10	10	Greenland halibut	Reinhardtius hippoglossoides	N	99-09	11,4	0,30	<0,25	<0,25	<0,25	9,4	<0,50	3,5	0,77
R20-1226-11	11	Mackerel	Scomber scombrus	n.a	30-40	26,5	0,28	<0,25	<0,25	<0,25	2,9	<0,50	0,88	0,72
R20-1226-12	12	Sea urchin	Echinoidea	MM	n.a	5,0	<0,25	<0,25	<0,25	<0,25	9,0	<0,50	<0,50	0,55
R20-1226-13	13	Cod liver	Gadus morhua	n.a	n.a	67,0	1,40	0,80	<0,25	0,58	83,0	2,13	24	3,08
R20-1226-14	14	Blue mussels	Mytilus edulis	MM	n.a	9,0	<0,25	<0,25	<0,25	<0,25	9,0	<0,50	<0,50	0,55
R20-1226-15	15	Blue whiting	Micromesistius poutassou	NW	30-40	1,2	<0,25	<0,25	<0,25	<0,25	8,0	<0,50	<0,50	0,55
R20-1226-16	16	Atlantic argentine	Argentina silus	SE	40-50	1,3	<0,25	<0,25	<0,25	<0,25	1,0	<0,50	<0,50	0,55
R20-1226-17	17	Deep-sea rosefish	Sebastes mentella	MS	40-50	2,3	<0,25	<0,25	<0,25	<0,25	1,3	<0,50	0,61	0,55
Table 2 (cont)	: Pesticides ir	Table 2 (cont): Pesticides in marine catches and one cod liver sample on	ne cod liver sample on wet weight											
				:	;	Lipid	Aldrin/		Octachloro		Endo-		trans-	
Sample code	Fish sample	Sample name	Latin name	Fishing	Size	content	dieldrin	Toxaphene	styrene	Endrin	s ul fane	Chlordane	Nonachlor	Mirex
	no.					%	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg
R20-1226-1	_	Rose fish	Sebastes marinus	NS	30-40	3	0,64	2,3	<0,1	<0,3	1,3	0,87	0,25	<0,1
R20-1226-2	2	Haddock	Melanogrammus aeglefinus	NE	40-49	9,0	0,25	2,0	<0,1	€,0>	1,3	0,70	<0,1	<0,1
R20-1226-3	3	Cod	Gadus morhua	N	50-59	9,0	0,25	2,0	<0,1	€,0>	1,3	0,70	<0,1	<0,1
R20-1226-4	4	Lemon sole	Microstomus kitt	NE	30-50	9,0	0,46	3,3	<0,1	9,0>	1,7	1,30	<0,1	<0,2
R20-1226-5	5	Thomy skate	Raja Amblyraja radiata	NE	40-60	9,0	0,25	2,0	<0,1	€,0>	1,3	0,70	<0,1	<0,1
R20-1226-6	9	Pollock	Pollachius virens	NW	02-09	8,0	0,25	2,0	<0,1	<0,3	1,3	0,70	<0,1	<0,1
R20-1226-7	7	Whiting	Merlangius merlangus	SE	40-50	9,0	0,25	2,0	<0,1	€,0>	1,3	0,70	<0,1	<0,1
R20-1226-8	∞	Atlantic wolffish	Anarhichas lupus	SE	30-40	9,0	0,25	2,0	<0,1	<0,3	1,3	0,70	<0,1	<0,1
R20-1226-9	6	Cod	Gadus morhua	N	02-09	8,0	0,25	2,0	<0,1	<0,3	1,3	0,70	<0,1	<0,1
R20-1226-10	10	Greenland halibut	Reinhardtius hippoglossoides	N	99-09	11,4	2,2	9,4	0,11	<0,3	1,3	2,47	2,8	0,17
R20-1226-11	11	Mackerel	Scomber scombrus	n.a	30-40	26,5	1,8	4,2	<0,1	€,0>	1,3	1,06	0,63	<0,1
R20-1226-12	12	Sea urchin	Echinoidea	NW	n.a	5,0	0,34	2,0	<0,1	0,3	1,3	0,70	<0,1	<0,1
R20-1226-13	13	Cod liver	Gadus morhua	n.a	n.a	67,0	21	75	1,07	1.96	1,3	21	27	1,10
R20-1226-14	14	Blue mussels	Mytilus edulis	NW	n.a	9,0	0,25	1,1	<0,1	0,3	1,3	0,70	<0,1	<0,1
R20-1226-15	15	Blue whiting	Micromesistius poutassou	NW	30-40	1,2	0,25	2,0	<0,1	€,0>	1,3	0,70	<0,1	<0,1
R20-1226-16	16	Atlantic argentine	Argentina silus	SE	40-50	1,3	0,25	1,1	<0,1	€,0>	1,3	0,70	0,12	<0,1
K20-1226-17	[]	Deep-sea rosetish	Sebastes mentella	SW	40-50	2,3	0,63	7,7	<0,1	<0,3	£,1	1,00	0,0	<0,1

g mg/kg mg/kg mg/kg 9 1,5 0,001 <0,002 0,10 3 9,7 0,001 <0,002 0,01 8 2,4 0,002 <0,002 0,00 15 13,0 0,002 <0,00 0,04 15 0,7 0,000 <0,00 0,04 16 0,000 <0,002 0,00 0,00 1,6 0,000 <0,002 0,00 0,00 1,6 0,000 <0,000 0,00 0,00 2 0,002 <0,002 0,00 0,00 6 5,5 0,002 <0,00 0,00 6 5,5 0,022 <0,01 0,00 6 3,6 0,304 <0,00 0,00 1 0,143 <0,002 0,00 1 2,5 0,004 <0,002 0,00 1 2,5 0,004 <0,002 0,00 1	Table 3: Trace	e elements in mari	one	cod liver sample in mg/kg wet weight	۲	ŠĄ	S	S.	Но	Ph
1 Rose fish Sebastes marinus 0,019 1,5 0,001 <0,002	Sample code	Fish sample no.	Sample name	Latin name		mg/kg	mg/kg	mg/kg	mg/kg	mg/kg
2 Haddock Melanogrammus aeglefinus 0,033 9,7 0,001 <0,002 0,001 3 Cod Gadus morhua 0,068 2,4 0,002 <0,002	R20-1226-1	1	Rose fish	Sebastes marinus	0,019	1,5	0,001	<0,002	0,10	<0,015
3 Cod Gadus morhua 0,068 2,4 0,002 <0,002 0,002 0,002 0,002 0,003 0,002 0,003 <th< td=""><td>R20-1226-2</td><td>2</td><td>Haddock</td><td>Melanogrammus aeglefinus</td><td>0,033</td><td>7,6</td><td>0,001</td><td><0,002</td><td>0,01</td><td><0,015</td></th<>	R20-1226-2	2	Haddock	Melanogrammus aeglefinus	0,033	7,6	0,001	<0,002	0,01	<0,015
4 Lemon sole Microstomus kitt 0,036 32,0 0,003 <0,002 0,005 5 Thomy skate Raja Amblyraja radiata <0,015	R20-1226-3	С	Cod	Gadus morhua	0,068	2,4		<0,002	0,02	<0,015
5 Thomy skate Raja Amblyraja radiata <0,015 13,0 0,002 <0,002 0,04 6 Pollock Pollachius virens <0,015	R20-1226-4	4	Lemon sole	Microstomus kitt	0,036	32,0	0,003	<0,002	0,05	<0,015
6 Pollock Pollachius virens <0,015 0,7 0,003 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,000 <0,0	R20-1226-5	5	Thomy skate	Raja Amblyraja radiata	<0,015	13,0		<0,002	0,04	<0,015
7 Whiting Merlangius merlangus 0,030 1,6 0,000 <0,000 <0,10 8 Atlantic wolffish Anarhichas lupus 0,027 6,8 0,002 <0,002	R20-1226-6	9	Pollock	Pollachius virens	<0,015	0,7	0,003	<0,002	0,03	<0,015
8 Atlantic wolffish Anarhichas lupus 0,027 6,8 0,002 <0,000 0,00	R20-1226-7	7	Whiting	Merlangius merlangus	0,030	1,6		<0,002	0,10	<0,015
9 Cod Gadus morhua 0,058 2,6 0,002 <0,002 0,044 10 Greenland halibut Reinhardtius hippoglossoides 0,017 2,7 0,001 <0,002	R20-1226-8	~	Atlantic wolffish	Anarhichas lupus	0,027	8,9	0,002	<0,002	0,10	<0,015
10 Greenland halibut Reinhardtius hippoglossoides 0,017 2,7 0,001 <0,002 0,006 0,006 0,002	R20-1226-9	6	Cod	Gadus morhua	0,058	2,6		<0,002	0,04	<0,02
11 Mackerel Scomber scombrus <0,06 5,5 0,022 <0,01 0,00 12 Sea urchin Echinoidea 0,22 6,6 0,136 <0,02	R20-1226-10	10	Greenland halibut	Reinhardtius hippoglossoides	0,017	2,7		<0,002	90,0	<0,015
12 Sea urchim Echinoidea 0,22 6,6 0,136 <0,002 <0,02 13 Cod liver Gadus morhua <0,06	R20-1226-11	111	Mackerel	Scomber scombrus	<0,0>	5,5	0,022	<0,01	90,0	<0,07
13 Cod liver Gadus morhua <0,06 3,6 0,304 <0,01 0,01 14 Blue mussels Mytilus edulis 0,19 1,0 0,143 <0,002	R20-1226-12	12	Sea urchin	Echinoidea	0,22	9,9		<0,002	<0,02	<0,02
14 Blue mus sels Mytilus edulis 0,19 1,0 0,143 <0,002 0,00 15 Blue whiting Micromesistius poutassou 0,01 3,7 0,004 <0,002	R20-1226-13	13	Cod liver	Gadus morhua	<0,0>	3,6	0,304	<0,01	0,01	<0,07
15 Blue whiting Micromesistius poutassou 0,01 3,7 0,004 <0,002 0,10 16 Atlantic argentine Argentina silus 0,01 2,5 0,002 <0,002	R20-1226-14	14	Blue mussels	Mytilus edulis	0,19	1,0	0,143	<0,002	0,00	<0,008
16 Atlantic argentine Argentina silus 0,01 2,5 0,002 <0,002 0,07 17 Deep-sea rosefish Sebastes mentella <0,015	R20-1226-15	15	Blue whiting	Micromesistius poutassou	0,01	3,7		<0,002	0,10	<0,015
17 Deep-sea rosefish Sebastes mentella <0,015 4,5 0,002 <0,008 0,18	R20-1226-16	16	Atlantic argentine	Argentina silus	0,01	2,5	0,002	<0,002	0,07	<0,015
	R20-1226-17	17	Deep-sea rosefish	Sebastes mentella	<0,015	4,5	0,002	<0,002	0,18	<0,015